

Technical Information

Double Sugar Agar, Russell

Product Code: DM 1057

Application: Double Sugar Agar, Russell is used for the differentiation of gram-negative enteric bacilli on the basis of their ability to ferment dextrose and lactose with or without gas formation.

Composition**		
Ingredients	Gms / Litre	
Peptic digest of animal tissue	2.500	
Casein enzymic hydrolysate Beef extract	7.500 3.000	
Lactose	10.000	
Dextrose	1.000	
Sodium chloride	5.000	
Phenol red	0.025	
Agar Final pH (at 25°C)	15.000 7.3±0.2	

Principle & Interpretation

Gram-negative bacilli belonging to family *Enterobacteriaceae* are most frequently isolates from clinical specimens. Final identification of the members of *Enterobacteriaceae* requires a battery of biochemical tests ⁽¹⁾. Double Sugar Agar, medium was originally formulated by Russell ⁽²⁾ using litmus indicator. It was later modified by Nichols ⁽³⁾ and Nichols and Wood ⁽⁴⁾ by replacing the litmus indicator with phenol red. This medium is used for differentiating gram-negative enteric bacilli especially the colon-typhoid-salmonellae-dysentery groups based on the fermentation of the double sugars incorporated namely, dextrose and lactose with or without gas formation. On incubation of inoculated tubed medium, acid production under aerobic condition (on the slant) and under anaerobic condition (in the butt) can be detected by the change in colour of the indicator. Phenol red is the pH indicator in the medium. Gaseous fermentation is indicated by splitting of the agar or by bubble formation in the butt. Organism like *Salmonella* Typhi capable of fermenting dextrose but not lactose will show an initial acid slant in short incubation of acids. Under anaerobic condition (in the butt), the same organism fails to revert the reaction and remains acidic. Peptic digest of animal tissue, casein enzymic hydrolysate and beef extract serve as sources of carbon, nitrogen, vitamins and other essential nutrients. Lactose and dextrose serve as sources of energy by being the fermentable carbohydrates. Phenol red is the pH indicator in the medium. Pure cultures are used to inoculate the tubed medium ⁽⁵⁾.

Methodology

Suspend 44.02 grams of powder media in 1000 ml distilled water. Shake well & heat to dissolve the medium completely. Dispense in tubes or as desired and sterilize by autoclaving at 118-121°C for 15 minutes. Allow the tubes to solidify in slanting position to form a generous butt.

Quality Control

Physical Appearance

Light yellow to pink homogeneous free flowing powder

Gelling

Firm, comparable with 1.5% Agar gel

Colour and Clarity of prepared medium

Red coloured, clear to slightly opalescent gel forms in tubes as slants

Reaction

Reaction of 4.4% w/v aqueous solution at 25°C. pH : 7.3±0.2





Bases / Media Supplements

pH range

7.10-7.50

Culture Response/Characteristics

DM1057: Culture characteristics observed after an incubation at 35-37⁰C for 18-40 hours.

Organism	Inoculum (CFU)	Growth	Slant	Butt	Gas
Enterobacter aerogenes ATCC 13048	50-100	Luxuriant	acidic reaction, yellowing of the medium	Acidic reaction yellowing of the medium	Positive reaction
Escherichia coli ATCC 25922	50-100	Luxuriant	acidic reaction, yellowing of the medium	Acidic reaction yellowing of the medium	Positive reaction
Proteus vulgaris ATCC 13315	50-100	Luxuriant	acidic reaction, yellowing of the medium	Acidic reaction yellowing of the medium	Positive reaction
Pseudomonas aeruginosa ATCC 27853	50-100	Luxuriant	alkaline reaction, red colour of the medium	Alkaline reaction red colour of the medium	Positive reaction
Salmonella Typhimurium ATCC 14028	50-100	Luxuriant	alkaline reaction, red colour of the medium	Acidic reaction yellowing of the medium	Positive reaction
Shigella dysenteriae ATCC 13313	50-100	Luxuriant	alkaline reaction, red colour of the medium	Acidic reaction yellowing of the medium	Positive reaction

Storage and Shelf Life

Dried Media: Store below 30°C in tightly closed container and use before expiry date as mentioned on the label. **Prepared Media:** 2-8⁰ in sealable plastic bags for 2-5 days.

Further Reading

1. Koneman E. W., Allen S. D., Janda W. M., Schreckenberger P. C., Winn W. C. Jr., 1992, Colour Atlas and Textbook of Diagnostic Microbiology, 4th Ed., J. B. Lippinccott Company

2. Russell F. F., 1911, J. Med. Res., 25:217.

3. Nichols H. J., 1921, J. Infect. Dis., 2982

4. Nichols H. J. and Woods C. B., 1922, J. Infect. Dis., 30, 320

5. MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. 1, Williams and Wilkins, Baltimore

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